

Role of quercetin in the prevention and treatment of diseases: Mini review

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Abstract. Quercetin is the most abundant flavonoid and one of the most important antioxidant of plant origin. The aim of the review was to describe quercetin and its role in the prevention and treatment of diseases. Articles were searched from internet databases using the following search words; quercetin, oxidative stress, quercetin and liver disease, quercetin and kidney disease, quercetin and hyperglycemia. The articles that met the selection criteria were used to describe quercetin and its role in the prevention and treatment of different diseases. The result showed that flavonoids are generally found at higher concentrations in outer layers of fruits and vegetables, onion has more quercetin than blackcurrants, broccoli, black grapes and apple. Quercetin and quercetin rich diets are used in the treatment and prevention of hyperglycemia, cardiovascular and kidney diseases, liver damage and nervous system disorders. In conclusion, quercetin is a naturally occurring flavonoid, more abundant in fruits and vegetables and are used in the treatment and prevention of many diseases.

Keywords: Quercetin; Flavonoid; Oxidative stress; Disease.

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Introduction

Quercetin (3,3',4',5,7-pentahydroxyflavone) is a flavonol, one of the six subclasses of flavonoid compounds, it is insoluble in cold water, poorly soluble in hot water but quite soluble in alcohol and lipids (Kelly, 2011). Quercetin is the most abundant flavonoid, widely

distributed throughout the plant kingdom and one of the most important antioxidant of plant origin (Crozier et al., 2009; Brüll et al., 2015). They occur naturally in fruits and vegetables including apples, berries, grapes, onions, brassica vegetables, shallots, tea and tomatoes (Kelly, 2011). Oxidative stress occur as a results of imbalance between

the generations of reactive oxygen species/free radicals and endogenous antioxidant systems. Free radicals and reactive oxygen species (ROS) are formed under normal physiological conditions but become deleterious when not eliminated by the endogenous systems. ROS are major sources of primary catalysts that initiate in vivo and in vitro oxidation and create oxidative stress which results in numerous diseases such as cancer (Kinnula and Crapo, 2004), Alzheimer's disease (Smith et al., 2000) Parkinson's disease (Bolton et al., 2000), alcohol induced liver disease (Arteel, 2003) and diabetes (Rajeshkumar, 2010). Oxygen derived free radicals such as superoxide anions, hydroxyl radicals and hydrogen peroxide are cytotoxic and can cause tissue injuries or increases the severity (Jainu and Devi, 2005). Excessive amount of ROS is harmful because they initiate bimolecular oxidation which causes oxidative stress that eventually results in malfunction of cells. In addition, oxidative stress causes inadvertent enzyme activation and oxidative damage to cellular system (Wiseman and Halliwell, 1996)

Antioxidants are molecule capable of slowing or preventing the oxidation of other molecules. Oxidation is a chemical reaction that transfers electrons from a substance to an oxidizing agent. Oxidation reactions can produce free radicals, which start chain reactions that damage cells. Antioxidants terminate these chain reactions by removing free radical intermediates, and inhibit other oxidation reactions by being oxidized themselves. Therefore, antioxidants are reducing agents, examples are thiols, quercetin, ascorbic acid or polyphenols (Kawsar et al., 2014). Although oxidation reactions are crucial for life, they can also be damaging; hence, plants and animals maintain complex systems of multiple types of antioxidants, such as glutathione, quercetin, vitamin C, and vitamin E as well as enzymes such as catalase, superoxide dismutase and

various peroxidases. Low levels of antioxidants, or inhibition of the antioxidant enzymes causes oxidative stress and may damage cells. Free radicals are chemical species associated with an odd number or unpaired electron. They are neutral, short lived, unstable and highly reactive substances. They are capable of attacking the healthy cells of the body, causing them to lose their structure and function. Cell damage caused by free radicals appears to be a major contributor to aging and diseases such as cancer, cardiovascular disease, liver diseases, diabetes mellitus, inflammation, renal failure, and brain disorders. In order to protect the cells and organs of the body against reactive oxygen species, humans have developed a highly sophisticated and complex antioxidant protection system that functions synergistically to neutralize free radicals (Kawsar et al., 2014). The aim of the review was to describe quercetin and its role in the prevention and treatment of diseases.

Materials and methods

Articles were searched from the Directory of Open Access Journals, Google Scholar, PubMed, Science Direct and Scopus using key words such as quercetin, oxidative stress, quercetin and liver disease, quercetin and kidney disease, quercetin and diabetes, quercetin and cardiovascular disease, quercetin and nervous system injury. The articles were selected and reviewed base on the following criteria:

- i. Articles on quercetin from different sources.
- ii. Articles that described the role of quercetin in the prevention and treatment of different diseases.
- iii. Articles on antioxidant and oxidative stress.

Fifty one (51) articles from different databases met the selection criteria and were selected for the review.

The articles were used to describe quercetin and its role in the prevention and treatment of different diseases.

Results

Quercetin

Quercetin is widely distributed in the plant kingdom and has a wide range of uses. Most of the dietary intake of quercetin-type flavonols is as quercetin glycosides. The most common are quercetin linked to one or two glucose molecules (quercetin glucosides) and quercetin linked to rutinose (quercetin rutinoside). The aglycone form of quercetin is not as abundant as the flavonol glycoside form. Two of the most important food sources of aglycone form of quercetin are onions and shallots, quercetin in shallot flesh is about 99.2% quercetin glucosides and 0.8% quercetin aglycone while the dry shallot skin consist of 83.3% quercetin aglycone and 16.7% quercetin glucosides (Wiczowski, et al., 2008). The flesh of onions contains mostly quercetin glucosides, with only trace amounts of quercetin aglycone while the skin and outermost layers of an onion have much more quercetin aglycone (Smith et al., 2003).

Flavonoids are classified into 13 different categories (Croft, 1998). Flavonoids serve as chemo-preventers in foods, they play a role as antioxidants preventing the rancidity development in lipids before consumption or during digestion processes. They increase intestinal transit time, protect intestinal microflora, can increase up take of some beneficial constituents from the diet, and reduce the level of food mutagens and carcinogens (Stavric et al., 1997). There are seven major flavonoid compounds in onions; Quercetin aglycone, quercetin monoglucoside, quercetin diglucoside, isorhamnetin (a methylether of quercetin), isorhamnetin monoglucoside, rutin and kaempferol (Park and Lee, 1996). Quercetin diglucoside and monoglucoside account for up to 93% of

the total flavonol content in onion (Lombard et al., 2002). Flavonoids are generally found at higher concentrations in outer layers of fruits and vegetables (Tsushida and Suzuki, 1996), therefore peeling results in their great loss. After homelike peeling red onions contained 79% of the original total content of quercetin-4'-glucoside and only 27% of the anthocyanins (Gennaro et al., 2002). Onion has more quercetin (300 mg/kg) than blackcurrants (40 mg/kg), broccoli, black grapes and apple (30 mg k/g) (Hollman and Arts, 2000). The total quercetin content in the dry onion skins is significantly higher than that in the edible parts. However the levels of quercetin glucosides in the dry outer skins are less than 10% of the levels in fleshy and partly dried scales. The probable mechanism is that quercetin is formed by deglucosidation of quercetin glucosides on the border between drying and dried brown areas on individual scales (Takahama & Hirota, 2000). About 90% of the total quercetin of each scale is confined to the epidermal tissue, and the rest in the storage tissue. The total content of quercetin is higher in the upper part of an onion as compared with the lower part (Trammell and Peterson, 1976).

Many studies have reported the preventive and therapeutic role of quercetin from different fruits and vegetables on various organs and tissues of the body in humans, cell lines and animal models. Quercetin was reported to protect the liver, kidney and heart of Wistar rats from doxorubicin induced toxicity (Jambhulkar et al., 2014).

Effect of quercetin on hyperglyceamia

Quercetin rich diet was reported to significantly reduce plasma glucose and blood glycated hemoglobin in diabetic mice compared to controls, it had no significant influence on plasma insulin level but significantly decrease the activities of small intestinal maltase activity (Kim et al., 2011). Therefore,

quercetin may be effective in controlling post prandial and fasting blood glucose levels in diabetic patients. Bakhshaeshi et al. (2012) reported the preventive effect of quercetin derived from *Allium cepa* on the liver of streptozotocin-induced diabetic rats, they showed that quercetin reduced the number of apoptotic cells in the liver suggesting its role in the prevention of liver damage that occur as a result of diabetes. Treatment of streptozotocin-induced diabetic rats with quercetin result in decreased blood glucose levels and reduction in the activities of ALT and AST as compared with untreated diabetic rats having high blood glucose levels due to impaired metabolism (Kilicarslam and Donmez, 2016).

Zhang et al. (2016) reported that administration of quercetin from *Toona sinensis* leaves (QTL) to diabetic mice significantly reduces the serum levels of glucose, insulin, total cholesterol, ALT, AST, triglycerides and low density lipoprotein-cholesterol compared with untreated diabetic mice. It further reduce oxidative stress as determined by lipid peroxidation and nitric oxide content as a result, decrease the rate of liver injury that occur in diabetic state. QTL also suppressed the diabetes-induced activation of p65/NF- κ B pathways, caspase-3 and caspase-9 levels in the liver as well as decrease in the levels of cellular organelle injury in the hepatocytes of diabetic rats. Quercetin significantly reduce fasting blood glucose and malondialdehyde (MDA) levels in diabetic rats compared to diabetic controls. The mRNA levels of HSP27, HSP70, HSF-1 and glucose-6-phosphate was also significantly decreased while the expression of glycokinase was significantly increased in response to quercetin (Hemmanti et al., 2018). This findings suggest that the therapeutic effect of quercetin could be through increase in transcript level of glucokinase that simultaneously decrease the expression of glucose-6-phosphate and stress protein (Hemmanti et al., 2018).

Effect of quercetin on liver disease

Bile duct obstruction in rats causes a decrease in hepatic and mitochondrial thiobarbituric acid reactive substances (TBARS), collagen concentration and fibrosis but administration of quercetin to the biliary obstructed rats resulted in reduced liver oxidative damage, ductal proliferation, fibrosis and a decrease in TBARS and collagen concentration, suggesting that quercetin can be used to preserve liver function in patients with biliary obstruction (Peres et al., 2000). Quercetin was reported to have protective effect on the liver of rats with Non-alcoholic steatohepatitis (NASH). There was a significant decrease in hepatic damage enzymes, lipoperoxidation, DNA damage and a lower micro-vesicular steatosis in NASH rats treated with 50 mg/kg quercetin compared to untreated NASH rats (Marcolin et al., 2013). Liposomal quercetin was reported to inhibit concavalin-A induced acute hepatitis and hepatic fibrosis. The probable mechanism was the ability of quercetin to modulate antinuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and transforming growth factor beta (TGF- β) production suggesting that liposomal quercetin can be a potent substance in the treatment of patients with liver damage and liver fibrosis (Wan et al., 2014).

Earlier study by Ashkani-Esfahani et al. (2016). Showed the protective effect of quercetin on thioacetamide-induced acute liver damage in Sprague-Dawley rats. Rats that received 350 mg/kg thioacetamide (TAA) plus 300 mg/kg quercetin intraperitoneally had a significant decrease in the level of ALT, AST and NH₄ with lower piecemeal necrosis and encephalopathy compared with rats treated with 350 mg/kg TAA only. The levels of AST, ALT, total bilirubin and triglycerides were reduced in alcohol-induced liver injured mice treated with

quercetin (Zhu et al., 2017). It also increased the activities of SOD, GPx and suppressed IL-1 β , IL-6, IL-10 and inducible nitric oxide synthase. Quercetin also suppressed the protein expression levels of B-cell lymphoma (Bcl)-2, caspase-3, poly ADP-ribose polymerase and signal transducer and activator of transcription (STAT) 3 phosphorylation, nuclear factor (NF)- κ B and protein kinase B (Akt) Phosphorylation levels in alcohol-induced liver injured mice. These results suggest that the role of quercetin against alcohol-induced liver injury was through the phosphoinositide 3-kinase/Akt/NF- κ B and STAT 3 pathway.

Wu et al. (2017) reported that quercetin was able to reduce bile duct ligation and carbon tetrachloride (CCl₄) liver cirrhosis in mice by inhibiting extracellular matrix formation and regulating matrix metalloproteinase (MMP)-9 and tissue inhibitor of metalloproteinase (TIMP)-1. The mechanism through which quercetin attenuate liver damage was by suppressing the TGF- β 1/smad signaling pathway and activating the P13k/Akt signaling pathway to inhibit autophagy. Quercetin prevents hepatic fibrosis by decreasing the rate of activation of hepatic stellate cell and inhibiting autophagy through the regulation of crosstalk between TGF- β 1/smad and P13/Akt pathway. Li et al. (2018) conducted a research to investigate the impact of quercetin on macrophages activation and polarization. BALB/c mice were induced liver fibrosis using CCl₄ for 8 weeks and concomitantly treated with 50 mg/kg quercetin. Quercetin was shown to ameliorate liver inflammation, fibrosis and inhibit HSCs activation. It also inhibited macrophage activation and M1 Polarization as well as decreased the mRNA expression of M1 macrophages markers such as TNF- α , IL-1 β , IL-6 and nitric oxide synthase 2. Quercetin reduces CCl₄-induced liver inflammation and fibrosis in mice through the inhibition of macrophages infiltration

and modulating M1 macrophages polarization via targeting Notch 1 pathway.

Effect of quercetin on kidney disease

Quercetin treatment significantly reduces diabetic nephropathy (DN) in hypercholesterolemic mice by decreasing serum levels of glucose and triglycerides and normalizing the glomerulosclerosis index and the kidney and body weight (Gomes et al., 2015). Quercetin administration ameliorates kidney fibrosis and macrophage accumulation in kidney with obstructive nephropathy (Ren et al., 2016).

Quercetin administration in rats with 5/6 nephrectomy was able to reduce the plasma level of malondialdehyde (MDA), increase glutathione peroxidase (GPx) activity and reduce the degree of fibrosis in kidney tissue as compared with untreated 5/6 nephrectomized rats (Loyal et al., 2017). Flavonoids prevent renal injuries associated with arterial hypertension by decreasing blood pressure and acting on the renal parenchyma. This is due to flavonoid interference with multiple signaling pathways known to produce renal injury and are independent of their blood pressure lowering effect (Vargas et al., 2018). Flavonoid consumption also prevents the adverse effect of high fat diet, type 1 and 2 diabetes on kidney function.

Effect of quercetin on gastric disorders

High dietary intake of quercetin is inversely associated to the risk of non-cardiac gastric adenocarcinoma, the protection appears to be particularly strong in women exposed to oxidative stress (Estrom et al., 2011). There was a significant reduction in the lesion index in the stomach of indomethacin-induced gastric ulcerated rats treated with 50 mg/kg of quercetin as compared with untreated rats. Also a significant increase in protein bound carbohydrate

complexes and nucleic acids was observed with significant decrease in volume of gastric juice, pepsin concentration and acid output in the rats treated with 50 mg/kg of quercetin as compared to the untreated ones (Shakeerabanu et al., 2011). This suggest that the gastro-protective effect of quercetin might be due to its cytoprotective nature. Pre-treatment of indomethacin-induced gastric ulcerated diabetic rats with quercetin caused a significant decrease in gastric ulcer index, MDA, IL-6, TNF- α and p53 levels with concomitant increase in SOD activity when compared with normal and diabetic rats treated with indomethacin alone (Khaleel et al., 2015).

Alkushi and Elsayy (2017) reported that quercetin can protect gastric mucosa against indomethacin-induced gastric ulceration than famotidine by the observed decrease in ulcer index, with mild inflammatory cell infiltration in the stomach of rats treated with 50 mg/kg of quercetin as compared with those of rats treated with 50 mg/kg famotidine.

Effect of quercetin on the nervous system disorders

Pretreatment of primary hippocampal cultures with quercetin significantly reduce A β (1-42)-induced cytotoxicity, protein oxidation lipid peroxidation and apoptosis in cultured neurons (Ansari et al., 2009). These findings suggest that quercetin may provide a promising approach for the prevention and treatment of neurodegenerative diseases. Quercetin glycosides rutin and isoquercetin were reported to have neuroprotective effect against 6-OHDA-induced rat pheochromocytoma (PC-12) cells by significantly increasing the activities of catalase, superoxide dismutase, Glutathione peroxidase and glutathione that were reduced by 6-OHDA in PC-12 cells (Magalingam et al., 2016). There was no significant difference in the activation of glutathione peroxidase and

glutathione enzymes between rutin and isoquercetin signifying that the two glycosides are equally important in protecting PC-12 cells against 6-OHDA toxicity. Both rutin and isoquercetin suppressed lipid peroxidation, MDA generation and prevented cell damage in 6-OHDA-induced neurotoxic PC-12 cells. Quercetin supplementation in diabetic and non-diabetic rats reported were to have neuroprotective effect by preventing glial and neuronal loss and the presence of reduced neuronal and glial body areas (Souza et al., 2017). This suggest that quercetin have the capacity of preventing cellular damages associated with long term diabetes mellitus. Yang et al. (2018) showed that quercetin (50 mg/kg) was able to suppress azodthymidine (AZT)-induced neuroinflammation by significantly inhibiting the expression of microglial and astrocytic markers induced by 100 mg/kg AZT in the mouse cortex, hippocampus and spinal cord. Co-administration of quercetin with AZT also attenuates the up-regulation of pro-inflammatory cytokines.

Effect of quercetin on cardiovascular disease

Philippine red *Allium cepa* was reported to decrease the serum level of LDL-cholesterol in Wistar rats (Pinedi and Calzada, 2013). They suggested the flavonoid content of the red *Allium cepa* to be responsible for the decrease in the serum level of LDL-cholesterol and that *Allium cepa* may decrease the risk of atherosclerosis. Onion skin quercetin at 162 mg/d was capable of decreasing blood pressure in hypertensive patients suggesting a cardio-protective effect of quercetin but without effect on the mechanistic parameters (Brüll et al., 2015). Quercetin was reported to reduce the force of contraction of porcine pulmonary arteries by limiting calcium release from the sarcoplasmic reticulum with no effect on voltage-operated channel (Banerjee, 2015). The transcriptional activity of nuclear factor

of transcription kappa B (NF-KB) and serum levels of interleukin 1 β and Tissue necrosis factor- α (TNF- α) were decreased by quercetin in patients with coronary artery disease as compared with untreated controls (Chekalina et al., 2018).

Quercetin was reported to significantly reduce the carcinogen activity of some cooked food mutagens including bay-region diol epoxides of benzo[a] pyrene and heterocyclic amines, these carcinogens reduce activation by cytochrome P-450 dependent mixed-function oxidases; quercetin inhibits these oxidases in vitro (Morris, 2001).

Conclusion

Quercetin is a naturally occurring flavonoid, it is more abundant in fruits and vegetables. Onion contain more quercetin than blackcurrants, broccoli, black grapes and apple. The total quercetin content in the dry onion skin is significantly higher than that in the edible parts. Quercetin is used in the treatment and prevention hyperglycemia, kidney disease, liver disease, cardiovascular disease and nervous system disorders.

Conflicts of interest

Authors declare that they have no conflict of interests.

References

- Alkushi, A. G. R.; Elsayy, N. A. M. Quercetin attenuates; indomethacin-induced acute gastric ulcer in rats. **Folia Morphologica**, v. 76, p. 252-261, 2017. <https://doi.org/10.5603/FM.a2016.0067>
- Ansari, M. A.; Abdul, H. M.; Joshi, G.; Opiia, W. O.; Butterfield, D. A. Protective effect of quercetin in primary neurons against A β (1-42): Relevance to Alzheimer's disease. **The Journal of Nutritional Biochemistry**, v. 20, p. 269-275, 2009. <https://doi.org/10.1016/j.jnutbio.2008.03.002>
- Arteel, G. E. Oxidants and antioxidants in alcohol-induced liver disease. **Gastroenterology**, v. 124, p. 778-790, 2003. <https://doi.org/10.1053/gast.2003.50087>
- Ashkani-Esfahani, S.; Bagheri, F.; Azarpira, N.; Elmira, E. E.; Emami, Y.; Hassanabadi, N.; Keshtkar, M. Protective effects of quercetin on thioacetamide-induced acute liver damage and its related biochemical and pathological alterations. **The Egyptian Journal of Internal Medicine**, v. 28, p. 123-127, 2016. <https://doi.org/10.4103/1110-7782.200965>
- Bakhshaeshi, M.; Khaki, A.; Fathiazad, F.; Khaki, A. A.; Ghadamkheir, E. Anti-oxidative role of quercetin derived from *Allium cepa* on aldehyde oxidase (OX-LDL) and hepatocytes apoptosis in streptozotocin-induced diabetic rat. **Asian Pacific Journal of Tropical Biomedicine**, v. 2, p. 528-531, 2012. [https://doi.org/10.1016/S2221-1691\(12\)60090-2](https://doi.org/10.1016/S2221-1691(12)60090-2)
- Banerjee, P. Reducing risk of cardiovascular disease: Exploring the effects of quercetin on the contractile force of vascular smooth muscle. **Journal of Purdue Undergraduate Research**, v. 5, p. 2-9, 2015. <https://doi.org/10.5703/jpur.05.1.01>
- Bolton, J. L.; Trush, M. A.; Penning, T. M.; Dryhurst, G.; Monks, T. J. Role of quinones in toxicology. **Chemical Research in Toxicology**, v. 13, p. 135-160, 2000. <https://doi.org/10.1021/tx9902082>
- Brüll, V.; Burak, C.; Stoffel-Wagner, B.; Wolfram, S.; Nickenig, G.; Müller, C.; Langguth, P.; Alteheld, B.; Fimmers, R.; Naaf, S.; Zimmermann, B. F.; Stehle, P.; Egert, S. Effects of a quercetin-rich onion skin extract on 24 h ambulatory blood pressure and endothelial function in overweight-to-obese patients with (pre-)hypertension: A randomised double-blinded placebo-controlled cross-over trial. **British Journal of Nutrition**, v. 114, p. 1263-1277, 2015. <https://doi.org/10.1017/S0007114515002950>
- Chekalina, N.; Burmak, Y.; Petrov, Y.; Borisova, Z.; Manusha, Y.; Kazakov, Y.; Kaidashev, I. Quercetin reduces the transcriptional activity of NF-kB in stable coronary artery disease. **Indian Heart Journal**, v. 70, p. 593-597, 2018. <https://doi.org/10.1016/j.ihj.2018.04.006>
- Croft, K. D. The chemistry and biological effects of flavonoids and phenolic acids. **Annals of the New York Academy of**

- Sciences**, v. 854, p. 435-442, 1998. <https://doi.org/10.1111/j.1749-6632.1998.tb09922.x>
- Crozier, A.; Jaganath, I. B.; Clifford, M. N. Dietary phenolic: Chemistry, bioavailability and effects on health. **Natural Product Reports**, v. 26, p. 1001-1043, 2009. <https://doi.org/10.1039/B802662A>
- Ekstrom, A. M.; Serafini, M.; Nyren, O.; Wolk, A.; Bosetti, C.; Bellocco, R. Dietary quercetin intake and risk of gastric cancer: Results from a population-based study in Sweden. **Annals of Oncology**, v. 22, p. 438-443, 2011. <https://doi.org/10.1093/annonc/mdq390>
- Gennaro, L.; Leonardi, C.; Esposito, F.; Salucci, M.; Maiani, G.; Quaglia, G.; Fogliano, V. Flavonoid and carbohydrate contents in Tropea red onions: Effects of homelike peeling and storage. **Journal of Agricultural and Food Chemistry**, v. 50, p. 1904-1910, 2002. <https://doi.org/10.1021/jf011102r>
- Gomes, I. B. S.; Porto, M. L.; Santos, M. C. L. F. S.; Campagnaro, B. P.; Gava, A. L.; Meyrelles, S. S.; Pereira, T. M.; Vasquez, E. C. The protective effects of oral low-dose quercetin on diabetic nephropathy in hypercholesterolemic mice. **Frontiers in Physiology**, v. 6, p. 1-8, 2015. <https://doi.org/10.3389/fphys.2015.00247>
- Hemmati, M.; Mostafavi, S. E.; Zarban, A.; Hoshya, R. Protective effects of quercetin on hyperglycemia and stress proteins expression in rats with streptozocin-induced diabetes. **Modern Care Journal**, v. 15, no. 2, e64964, 2018. <https://doi.org/10.5812/modernc.64964>
- Hollman, P. C. H.; Arts, I. C. W. Flavonols, flavones and flavanols: Nature, occurrence and dietary burden. **Journal of the Science of Food and Agriculture**, v. 80, p. 1081-1093, 2000. [https://doi.org/10.1002/\(SICI\)1097-0010\(20000515\)80:7<1081::AID-JSFA566>3.0.CO;2-G](https://doi.org/10.1002/(SICI)1097-0010(20000515)80:7<1081::AID-JSFA566>3.0.CO;2-G)
- Jainu, M.; Devi, C. S. *In vitro* and *in vivo* evaluation of free radical scavenging potential of *Cissus quadrangularis*. **Pharmaceutical Resiew**, v. 43, p. 95-99, 2005. <https://doi.org/10.1080/13880200500406636>
- Jambhulkar, S.; Deshiredy, S.; Jestadi, D. B.; Periyasamy, L. Quercetin attenuating doxorubicin induced hepatic, cardiac and renal toxicity in male albino Wistar rats. **American Journal of Phytomedicine and Clinical Therapeutics**, v. 2, p. 985-1004, 2014.
- Kawsar, M. H.; Raihana, R.; Sultana, T.; Sohel, D.; Sohaily, S. I. *In-vitro* and *in-vivo* models for antioxidant activity evaluation: A review. **Journal SUB**, v. 5, p. 21-31, 2014.
- Kelly, G. S. Quercetin. **Alternative Medicine Review**, v. 16, p. 172-194, 2011.
- Khaleel, E. F.; Mostafa, D. G.; Abdel-Aleem, G. A. Gastroprotective effect of flavonoid quercetin and coenzyme Q10 in indomethacin-induced gastric ulcers in normal and diabetic rats. **OSR Journal of Dental and Medical Sciences**, v. 14, p. 58-71, 2015.
- Kilicarslan, G.; Donmez, N. The effects of quercetin on antioxidant system and some blood parameters in experimental diabetic rats. **Bulletin of Environment, Pharmacology and Life Sciences**, v. 5, p. 28-32, 2016.
- Kim, J.; Kang, M.; Choi, H.; Jeong, S.; Lee, Y.; Kim, J. Quercetin attenuates fasting and postprandial hyperglycemia in animal models of diabetes mellitus. **Nutrition Research and Practice**, v. 5, p. 107-111, 2011. <https://doi.org/10.4162/nrp.2011.5.2.107>
- Kinnula, V. L.; Crapo, J. D. Superoxide dismutases in malignant cells and human tumors. **Free Radical Biology and Medicine**, v. 36, p. 718-744, 2004. <https://doi.org/10.1016/j.freeradbiomed.2003.12.010>
- Layal, K.; Perdhana, I. S.; Louisa, M.; Estuningtyas, A.; Soetikno, V. The effects of quercetin on oxidative stress and fibrosis markers in chronic kidney disease rat model. **Medical Journal of Indonesia**, v. 26, p. 169-177, 2017. <https://doi.org/10.13181/mji.v26i3.1462>
- Li, X.; Jin, Q.; Yao, Q.; Xu, B.; Li, L.; Zhang, S.; Tu, C. The flavonoid quercetin ameliorates liver inflammation and fibrosis by regulating hepatic macrophages activation and polarization in mice. **Frontiers in Pharmacology**, v. 9, p. 1-14, 2018. <https://doi.org/10.3389/fphar.2018.00072>
- Lombard, K. A.; Geoffriau, E.; Peffley, E. Flavonoid quantification in onion by spectrophotometric and high performance liquid chromatography analysis. **HortScience**, v. 37, p. 682-685, 2002.

- Magalingam, K. B.; Radhakrishnan, A.; Haleagrahara, N. Protective effects of quercetin glycosides, rutin, and isoquercetrin against 6-hydroxydopamine (6-OHDA)-induced neurotoxicity in rat pheochromocytoma (PC-12) cells. **International Journal of Immunopathology and Pharmacology**, v. 29, p. 30-39, 2016. <https://doi.org/10.1177/0394632015613039>
- Marcolin, E.; Forgiarini, L. F.; Rodrigues, G.; Tieppo, J.; Borghetti, G. S.; Bassani, V. L.; Picada, J. N.; Marroni, N. P. Quercetin decreases liver damage in mice with non-alcoholic steatohepatitis. **Basic & Clinical Pharmacology & Toxicology**, v. 112, p. 385-391, 2013. <https://doi.org/10.1111/bcpt.12049>
- Morris, J. L. **Composition and flavonoid levels in Onion (*Allium cepa*) grown in hydroponies in greenhouses and growth chambers**. Texas: Faculty of Texas Technical University, 2001. (M. Sc. Thesis).
- Park, Y. K.; Lee, C. Y. Identification of isorhamnetin 4'-glucoside in onions. **Journal of Agricultural and Food Chemistry**, v. 44, p. 34-36, 1996. <https://doi.org/10.1021/jf950310e>
- Peres, W.; Tuñón, M. J.; Collado, P. S.; Herrmann, S.; Marroni, N.; González-Gallego, J. The flavonoid quercetin ameliorates liver damage in rats with biliary obstruction. **Journal of Hepatology**, v. 33, p. 742-750, 2000. [https://doi.org/10.1016/S0168-8278\(00\)80305-0](https://doi.org/10.1016/S0168-8278(00)80305-0)
- Pineda, M. R. B.; Calzada, G. Effect of Philippine red *Allium cepa* Lin. (Sibuyas na Pula) on serum LDL-cholesterol level. **International Journal of Medical and Biomedical Sciences**, v. 1, p. 14-20, 2013.
- Rajeshkumar, D. **Evaluation of antioxidant property and toxicological assessment of *Polyalthia longifolia* var. *Pendula* leaf**. Saurashtra: Saurashtra University, 2010. (Thesis of doctorate).
- Ren, J.; Li, J.; Liu, X.; Feng, Y.; Gui, Y.; Yang, J.; He, W.; Daia, C. Quercetin inhibits fibroblast activation and kidney fibrosis involving the suppression of mammalian target of Rapamycin and β -catenin signaling. **Scientific Reports**, v. 6, 23968, 2016.
- Shakeerabanu, M.; Sujatha, K.; Rajneesh, C. P.; Manimaran, A. The defensive effect of quercetin on indomethacin induced gastric damage in rats. **Advances in Biological Research**, v. 5, p. 64-70, 2011.
- Smith, C.; Lombard, K. A.; Peffley, E. B.; Weixin, L. W. Genetic analysis of quercetin in onion (*Allium cepa* L.) 'Lady Raider'. **Texas Journal of Agriculture and Natural Resources**, v. 16, p.24-28, 2003.
- Smith, M. A.; Rottkamp, C. A.; Nunomura, A.; Raina, A. K. Perry G. Oxidative stress in Alzheimer's disease. **Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease**, v. 1502, p. 139-144, 2000. [https://doi.org/10.1016/S0925-4439\(00\)00040-5](https://doi.org/10.1016/S0925-4439(00)00040-5)
- Souza, S. R. G.; Miranda Neto, M. H.; Colombo, J. V.; Perles, M.; Frez, F. C. V.; Zignani, I.; Ramalho, F. V.; Hermes-Uliana, C.; Bossolani, G. D. P.; Zanoni, J. N.. Antioxidant effects of the quercetin in the jejunal myenteric innervation of diabetic rats. **Frontiers in Medicine**, v. 4, p. 1-8, 2017. <https://doi.org/10.3389/fmed.2017.00008>
- Stavric, B.; Lau, B. P.-Y.; Matula, T. I.; Klassen, R.; Lewis, D.; Downie, R. H. Mutagenic heterocyclic aromatic amines (HAAs) in 'processed food flavour' samples. **Food and Chemical Toxicology**, v. 35, p. 185-197, 1997. [https://doi.org/10.1016/S0278-6915\(96\)00119-6](https://doi.org/10.1016/S0278-6915(96)00119-6)
- Takahama, U.; Hirota, S. Deglucosidation of quercetin glucosides to the aglycone and formation of antifungal agents by peroxidase-dependent oxidation of quercetin on browning of onion scales. **Plant & Cell Physiology**, v. 41, p. 1021-1029, 2000. <https://doi.org/10.1093/pcp/pcd025>
- Trammell, K. W.; Peterson, C. E. Quantitative differences in the flavonol content of yellow onion, *Allium cepa* L. **Journal of the American Society for Horticultural Science**, v. 101, p. 205-207, 1976.
- Tsushida, T.; Suzuki, M. Content of flavonol glucosides and some properties of enzymes metabolizing the glucosides in onion: Flavonoids in fruits and vegetables, part II. **Nippon Shokuhin Kagaku Kogaku Kaishi**, v. 43, p. 642-649, 1996. <https://doi.org/10.3136/nskkk.43.642>
- Vargas, F.; Romecín, P.; García-Guillén, A. I.; Wangesteen, R.; Vargas-Tendero, P.; Paredes, M. D.; Atucha, N. M.; García-Estañ, J. Flavonoids in kidney health and disease. **Frontiers in Physiology**, v. 9, p. 1-12, 2018. <https://doi.org/10.3389/fphys.2018.00394>

- Wan, Y.; Tang, M. H.; Chen, X. C.; Chen, L. J.; Wei, Y. Q.; Wang, Y. S. Inhibitory effect of liposomal quercetin on acute hepatitis and hepatic fibrosis induced by concanavalin A. **Brazilian Journal of Medical and Biological Research**, v. 47, p. 655-661, 2014. <https://doi.org/10.1590/1414-431X20143704>
- Wiczkowski, W.; Romaszko, J.; Bucinski, A. Z. Quercetin from shallots (*Allium cepa* L. var. *aggregatum*) is more bioavailable than its glucosides. **The Journal of Nutrition**, v. 138, p. 885-888, 2008. <https://doi.org/10.1093/jn/138.5.885>
- Wiseman, H.; Halliwell, B. Damage to DNA by reactive oxygen and nitrogen species: Role of inflammatory disease and progression to cancer. **Biochemical Journal**, v. 313, p. 17-29, 1996. <https://doi.org/10.1042/bj3130017>
- Wu, L.; Zhang, Q.; Mo, W.; Feng, J.; Li, S.; Li, J.; Liu, T.; Xu, S.; Wang, W.; Lu, X.; Yu, Q.; Chen, K.; Xia, Y.; Lu, J.; Xu, L.; Zhou, Y.; Fan, X.; Guo, C. Quercetin prevents hepatic fibrosis by inhibiting hepatic stellate cell activation and reducing autophagy via the TGF- β 1/Smads and PI3K/Akt pathways. **Scientific Reports**, v. 7, 92-89, 2017. <https://doi.org/10.1038/s41598-017-09673-5>
- Yang, Y.; Liu, X.; Wu, T.; Zhang, W.; Shu, J.; He, Y.; Tang, S. Quercetin attenuates AZT-induced neuroinflammation in the CNS. **Scientific Reports**, v. 8, no. 1, 6194, 2018. <https://doi.org/10.1038/s41598-018-24618-2>
- Zhang, Y.; Dong, H.; Wang, M.; Zhang, J. Quercetin Isolated from *Toona sinensis* leaves attenuates hyperglycemia and protects hepatocytes in high-carbohydrate/high-fat diet and alloxan induced experimental diabetic mice. **Journal of Diabetes Resesearch**, v. 2016, Article ID 8492780, 2016. <https://doi.org/10.1155/2016/8492780>
- Zhu, M.; Zhou, X.; Zhao, J. Quercetin prevents alcohol-induced liver injury through targeting of PI3K/Akt/nuclear factor- κ B and STAT3 signaling pathway. **Experimental Therapeutic Medicine**, v. 14, p. 6169-6175, 2017. <https://doi.org/10.3892/etm.2017.5329>

